

Original Research

Microplastic Pollution in Shoreline Sediments of Selected Rivers of Mizoram, North-East India – A Baseline Assessment and Preliminary Report

Joseph Vanlalsawma Sailo, Malsawmhriatzuala Jeremy and Laldinsangi Chawngte†

Department of Life Sciences, Pachhunga University College, Mizoram University (A Central University), Aizawl, 796001, Mizoram, India

†Corresponding author: Dr. Laldinsangi Chawngte; laldinsangi.c@gmail.com; laldinsangi@pucollege.edu.in

ORCID: <https://orcid.org/0000-0002-9636-9341>

Key Words	Microplastics, sediments, pollution, Himalayas, Mizoram, river
DOI	https://doi.org/10.46488/NEPT.2026.v25i03.B4421 (DOI will be active only after the final publication of the paper)
Citation for the Paper	Sailo, J.V., Jeremy, M. and Chawngte, L., 2026. Microplastic pollution in shoreline sediments of selected rivers of Mizoram, north-east India – A baseline assessment and preliminary report. <i>Nature Environment and Pollution Technology</i> , 25(3), B4421. https://doi.org/10.46488/NEPT.2026.v25i03.B4421

Abstract: Microplastics have emerged as another dimension of hazardous and persistent pollutants afflicting every corner of the globe, even permeating into critical biodiversity hotspots. This study aims to serve as a preliminary report on the occurrence and assessment of MP pollution in shore sediments of four selected rivers in the state of Mizoram, which lies in one of the sensitive biodiversity hotspots in India, and is also a part of the eastern Himalayan range. There have been minimal studies on microplastic pollution in this region despite their critical ecological location. Four freshwater rivers, namely Chite, Tlawng, Serlui-A and Tuirial were selected for the study due to their close proximity to urban settlements, and have been subjected to rampant plastic pollution. The study revealed that MPs were detected from all sampling sites, and abundance was highest in Chite, which runs through parts of Aizawl city. Particles within the size range of 0.15-0.25 mm were found to be the highest in numbers in all sampling locations while MPs between 3-5 mm were the least in number. The shapes of MPs were varied, constituting fragments, fibers, pellets and spheres, with fibers (40%) being the dominant shape overall. Polyethylene was found to be the dominant polymer type among particles analyzed using FTIR spectroscopy. This study contributes to the imperative assessment of MP pollution in river sediments of the eastern Himalayan region of India, as comprehensive research in this regard is still lacking.

1. INTRODUCTION

Plastics are an indispensable part of modern life, and they are used extensively in all parts of the globe. However, their indiscriminate use and improper disposal, along with inadequate and futile recycling efforts have contributed to a massive scale of plastic pollution in all types of environments, both terrestrial and aquatic (Rhodes 2018, Bajt 2021). Consequently, plastic pollution has yielded another dimension of hazardous pollutants in the form of microplastics (MPs) which are classified as miniscule plastic objects that range in size between 0.1 and 5mm (Desforges et al. 2014). These particles have been categorized as pollutants of emerging concern, and their detection have been reported in terrestrial and aquatic ecosystems, in the air, and even inside the tissues of living organisms, including humans (Smith et al. 2018, Goyal et al. 2023, Zhao et al. 2023). Aquatic environments in particular are highly susceptible to MP pollution, due to surface water run-offs that carry MP particles from anthropogenic sources such as industries, agricultural fields, waste dumping grounds and even personal care products (Rhodes 2018, Koelmans et al. 2019). Plastics are manufactured with the addition of various chemicals additives to enhance the quality and performance of the products such as dyes, lubricants, plasticizers, flame retardants, heat stabilizers and anti-microbials (Smith et al. 2018, Sendra et al. 2021) and several studies have suggested that there is a high possibility of these substances leaching into their immediate environment via ingested MPs, leading to deleterious effects in living systems that ingest them directly or indirectly (Bridson et al. 2023, Gulizia et al. 2023). MP pollution in aquatic environments, both freshwater and marine, are particularly concerning, due to their positions as critical entry points into the food chain, that may lead to bioaccumulation and subsequently affect a wide range of organisms, including humans. Numerous studies have reported such findings (Cole et al. 2014; Van Cauwenberghe & Janssen 2014, Krause et al. 2021, Patil et al. 2022, Gunaalan et al. 2023). MPs of different size ranges have been detected in both freshwater and marine environments all across the globe, in different continents and such identifications have been reviewed extensively (Xu et al. 2020, Vivekanand et al. 2021; Wang et al. 2022). MPs have been classified into various categories depending on their physical characteristics such as fragments, pellets, filaments, and granules. Chemically, MPs generally constitute monomers of polyethylene (PE), polyethylene terephthalate (PET), polypropylene (PP), polystyrene (PS), and polyvinyl chloride (PVC), high-and low-density polyethylene (HD/LD-PE), etc. (Anbumani & Kakkar 2018).

MPs in rivers accumulate through numerous sources including household hygiene items that are used for bathing and cleansing clothes, plastic wastes, and fabric wastes (Ghosh et al. 2023). Washing synthetic fabrics can result in generation of significant amounts of microplastic fibres (MFs) in wastewater which can then enter river systems (McCormick et al. 2016). Additionally, extensive use of fishing nets and lines that are left in the water, angling and other techniques used to catch fish (Ghanbari et al. 2022, Karthikeyan et al. 2022) also contribute to MP contamination. Other prominent sources include vehicle tires, city dust and road markings (Ramaremisa et al. 2022). Several freshwater systems serve as major routes for improperly managed plastic debris to enter the oceans. It has been reported that roughly 80% of plastic debris enter the oceans from land-based waste via streams and rivers, and are further weathered by the natural elements into smaller particles such as MPs (Wang et al. 2024).

This issue of MP pollution has gained considerable concern and traction especially within the last decade due to their persistence and ubiquity in all corners of the globe. Considering the possible havoc they reportedly wreak on living organisms that are exposed to these particles, detection and analysis of the extent of pollution in different ecosystems are urgently warranted, especially in areas such as biodiversity hotspots where their effects could be disastrous. One such example is the state of Mizoram in the eastern Himalayan region of India. The state is a small land locked region in the north-east corner of the country that lies within one of the recognized biodiversity hotspots in the world (Pautu et al. 2023) and boasts of endemic and new species of plants and animals being discovered constantly. Although plastic pollution is a persistent problem that has devastated several river systems within the state as well as other rivers in the Himalayan region, there is still an acute lack of studies pertaining to the extent and impacts of MP presence and pollution in the rivers of Mizoram. Most studies focusing on MP pollution in India have been limited to the sediments and waters of the eastern and western coastal regions, with minimal reports from inland water bodies in the Himalayan regions (Tsering et al. 2021).

Aizawl, the capital of Mizoram, is a densely populated city situated in the northeast corner of India. Being the largest and most heavily populated city in the state, it has also seen a rapid increase in plastic pollution especially within the last two decades, and improper disposal of garbage has considerably polluted several rivers and streams that are close to the urban settlements. One study by Singh et al. (2023) quantifying plastic waste generation in Aizawl city reported plastic consumption of 13.06 g/capita/day, with plastic packaging contributing the most, followed by carry bags in residential, commercial and dumping areas. This study also highlights the frequent improper disposal of these plastic wastes that choke drains, and eventually make their way into streams and rivers. An unfortunate example is the Chite river which runs through certain areas of Aizawl city, and indiscriminate dumping of plastic garbage has severely decimated this river. Although there is evident plastic pollution in the rivers that are in close proximity to Aizawl, there has been no comprehensive study on the detection and extent of MP pollution. Determining the degree of pollution in these freshwater rivers is crucial, since they are critical lifelines for the people. One of these rivers selected for the assessment, Serlui-A, is from where drinking water for the municipality of Aizawl is sourced. Another river selected for the study, Tuirial, also lies close to the Aizawl municipal garbage dumping ground, rendering it extremely vulnerable to MP runoff. Since there have been minimal investigations to evaluate plastic and MP occurrence in the state, this study, therefore, aims to address this existing data gap for the first time for this region. The investigation serves to provide a baseline assessment of MP pollution in riverbank sediments of selected rivers that are close to residential and commercial settlements, namely Chite, Tuirial, Tlawng, and Serlui-A during late monsoon (September, 2023). However, since this study was conducted as a preliminary baseline assessment (as no prior studies have been done in this regard) and no funding was received for the work, only one time point (i.e., late monsoon) has been selected to gauge the extent of MP pollution in riverbank sediments.

2. MATERIALS AND METHODS

Site of study:

This study was conducted in four selected rivers of Mizoram, north-east India, which lies in the Eastern Himalayan region (Fig. 1). The four rivers namely, Chite, Tlawng, Tuirial, and Serlui-A were selected based on their proximity to residential and commercial areas as well as their susceptibility to waste-runoff from urban areas. Detailed information regarding the sampling sites is given in the Table 1. Their location is cause for higher susceptibility to plastic pollution as a result of high population density, as well as prevalent cases of improper garbage disposal in and around the municipal areas. Chite flows into Tuirial river, and Serlui-A is also a tributary of Tlawng. Sample collection sites are shown in Fig. 2.

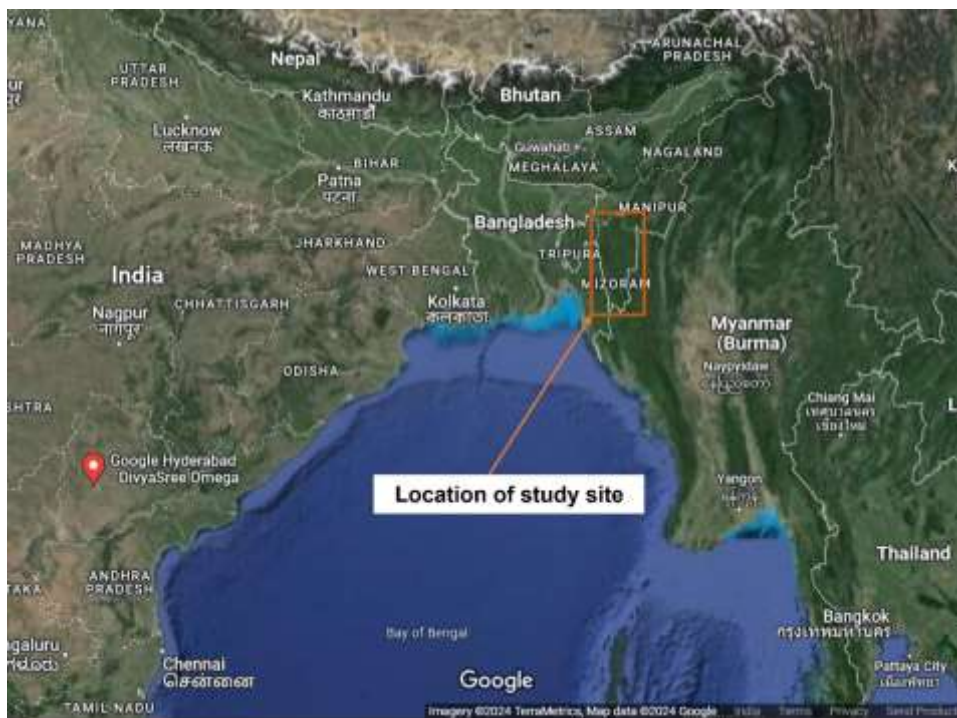


Fig. 1: Google Map showing the geographical location of the study site i.e., Mizoram, north-east India

Table 1: Location details of the sampling sites

Sl.No	River	Latitude and Longitude	Population Degree	River Section
1.	Chite	23°43'59.2" N, 92°44'17.5" E	Urban	Upstream
2.	Tlawng	23°42'43.46" N, 92°39'51.17" E	Rural	Upstream
3.	Serlui A	23°54'00" N, 92°36'36" E	Rural	Downstream
4.	Tuirial	23°43'07" N, 92°47'58" E	Semi-urban	Downstream



Fig. 2: Collection sites of shore sediment samples. (A) Google Map showing locations of study sites in Tlawng and Serlui-A. (B, C) Sample collection sites in Tlawng and Serlui-A respectively (D, E) Google Map of Chite and Tuirial respectively. (F, G) Sample collection location in Tuirial and Chite respectively.

Collection of samples:

Sample collection was performed as per the method stated by Tibbetts et al. (2018). In brief, sediment samples were collected from a selected site in the banks of the four selected rivers and samples were all collected within the month of September, 2023 i.e., the monsoon season when there is an increased rate of rainfall and water flow in rivers.

At all collection sites from each riverbank, samples from four different spots at depths of 10 cm were collected with a small steel shovel into 250 ml glass beakers, which resulted in a 1 kg bulk sample of sediment ($n=3$). Two samples were collected with a distance of approximately 3 feet between collection sites and the other two were collected 10 feet from the previous two and again spaced approximately 3 feet from each other (Tibbetts et al. 2018). The sediment samples from each river were stored in steel containers prewashed with distilled water and then brought into the laboratory for further treatments. The 1 kg mass sediments from each river were covered using aluminum foil right after collection to prevent plastic contamination. The materials that were used for collecting sediments were cleaned with filtered or distilled water to wash off other contaminants and plastics (Sarijan et al. 2018).

Sample processing:

The sediments were sieved into five different fractions using brass test sieves having mesh sizes of 5 mm, 3 mm, 1 mm, 0.5 mm, 0.25 mm, and 0.15 mm (Manikarn test sieve, MSWLABS, India). Hence the size range for each fraction is 0.15-0.25 mm, 0.25-0.5 mm, 0.5-1 mm, 1-3 mm and 3-5 mm. Larger stone particles and other kinds

of identifiable organic debris visibly larger than 5 mm were identified and manually removed. The sediments were wet sieved with filtered water to speed up the process and to separate the sediments and MPs into their respective fraction sizes. The sieved sediments with 5 different size fractions were then transferred to separate 250 ml beakers, and all beakers were marked with their river source. All beakers were covered with aluminum foil to prevent contamination by plastics from surroundings. Excess water contained in the sediments were carefully removed with filter paper. Alkali digestion using 10% KOH was done to remove any remaining organic matter, at a volume ratio of 1:3 (Saad et al. 2022). Sediment samples were dried in a hot air oven for 45 hours at 50°C. After drying, sediments for each size fraction were taken in a marked beaker for the density separation.

Density separation using ZnCl₂:

Zinc chloride (ZnCl₂) solution was used for density separation of MPs. The solution of 1.5 g/cm³ density was prepared by dissolving 1500 g of ZnCl₂ into 1 L (final volume) of distilled water in a laminar flow hood to avoid air-contamination. This was then poured into the samples and rigorously stirred for 40 seconds. Samples were then made to sit untouched until all the sediments settled into the bottom. The supernatant containing floating MPs were then allowed to overflow into a petri dish by the addition of fresh ZnCl₂. To improve extraction efficiency and minimize particle loss, two consecutive density separation cycles were conducted for each sediment sample, and the supernatants from both cycles were then combined prior to analysis. The particles separated via this method were then transferred to a clean petri dish, covered and were oven-dried at 37°C for 48 hours. The selected density was considered sufficient to float common polymers such as PE, PP, PS and a substantial proportion of PET and PA particles, while allowing efficient separation from the mineral sediment matrix.

Morphological identification, quantification and chemical analysis of MPs:

The isolated particles from all size fractions were visually identified based on their physical characteristics and quantified carefully under a light microscope (Leica S9i, Leica Microsystems) as proposed in various literature. The particles were sorted mainly on their sizes as well as two main shapes i.e., fragments and fibers. Since the accuracy and effectiveness of visual identification in extremely small MPs can be unreliable (Balestra & Bellopede, 2022), particles smaller than 0.15 mm were not taken into consideration and also pose difficult to handle. Quantification of MPs was done by counting the number of identifiable particles for each size fraction. Replicate samples were treated as within-site subsamples since collection was done only at one location per river, and statistical comparisons reflect variability among sampling sites rather than independent river-scale units.

In addition, several MP particles of different shapes were selected randomly, and their surface characteristics were analyzed using a scanning electron microscope (SEM; Hitachi TM 4000 Plus). MPs were placed on a carbon tape on a metal stub and then coated with gold using ion sputter MC1000 for 15 minutes and viewed under Scanning Electron Microscope (HITACHI TM4000Plus) at 15 kV.

To determine polymer composition of isolated MPs, 20 particles (5 from each sampling location) from the 0.5 to 0.1 mm range were randomly selected for Fourier transform infrared spectroscopy (FTIR) analysis. Functional group identification was done using a Bruker 3000 Hyperion FTIR spectrometer (Germany) between 400 and 4000 cm^{-1} and samples were recorded on OPUS software. Spectra analysis was done using comparison to frequently applied polymer libraries that have been used in other studies (Karing et al. 2023). Five MPs could not be conclusively identified as a result of weak spectra.

Contamination Control

To prevent secondary plastic contamination via laboratory equipment, sample collection, processing and separation were done using only metal tools or glassware, which were cleaned with filtered water prior to usage. All glassware used were covered in aluminum foil prior to sample loading. Samples collected were covered with aluminum foil during transportation to keep out any foreign particles. Contamination was avoided in the laboratory by use of cotton lab coats and nitrile gloves, and work spaces were cleaned thoroughly with ethanol. Use of plasticware was avoided during any handling of the samples. However, procedural laboratory blanks, field blanks, and airborne fallout controls were not included in the present study, and therefore no quantitative contamination correction was applied to the results reported for MP abundances.

Statistical Analysis

MP abundance was quantified using replicate samples ($n=3$) for all four rivers across five particle-size ranges. The statistical analyses were performed using GraphPad Prism and expressed as mean \pm SD (standard deviation). The conformity of variables with the normal distribution and homogeneity of variance were examined using Shapiro–Wilk normality test and Levene tests, respectively. All the quantitative data were analyzed using One-way analysis of variance (ANOVA) followed by Tukey test with significance level set at $p < 0.05$. Descriptive statistics (mean, SD, and 95% CI) were calculated for each river \times size-class combination. Confidence intervals were computed using the formula:

$$CI_{95} = t_{0.975,df-2} \times \frac{SD}{\sqrt{3}}$$

Total MP abundance per river and overall pooled statistics were calculated from summed replicate counts.

3. RESULTS

From the analysis, MPs of assorted sizes were detected in the sediment samples of all rivers under study, with varying quantities found in each river. The abundance of MPs of all size ranges was determined as particles per kilogram of sediment.

3.1 MP abundance:

In the study, MP particles isolated from each selected site at the four rivers were found to be varying in quantity depending on their size fractions, with a mean abundance of 238.17 ± 46.95 particles/kg of sediment (average \pm standard deviation) with a 95% CI of ± 29.80 particles, which highlights a moderate variability among sampling sites overall. The number and abundance of MPs per sample are given in Fig. 3.

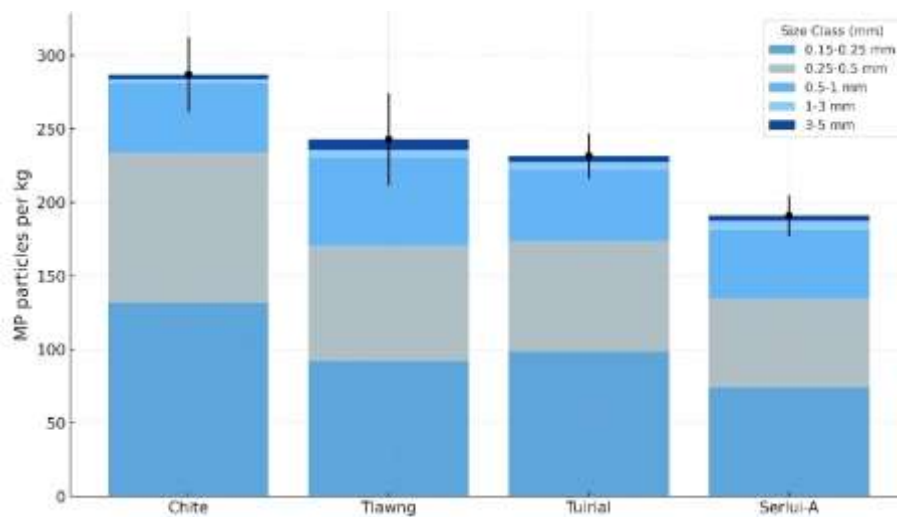


Fig. 3: Graphical representation of the number of MPs in Chite, Tlawng, Serlui-A, and Tuorial.

Mean abundances of MPs from each site of the four rivers differed considerably, with the highest number of MPs detected in Chite sediments (287 ± 43.41 particles), which could be attributed to the close distance to urban areas and the excessive levels of plastic pollution. Studies have also shown a positive correlation between proximity of freshwater bodies to urban areas with elevated levels of MPs (Wang et al. 2017, Ramaremissa et al. 2022). This was followed by Tlawng (243 ± 51.12) and Tuorial (231.67 ± 16.20). Serlui-A (191 ± 18.52) was found to have the lowest number of MP numbers which could be inferred as a result of its furthest distance from Aizawl compared to the other rivers. Although these differences show a clear numerical gradient, pairwise statistical analysis revealed that only the difference between Chite and Serlui-A was statistically significant (Welch's t-test: $t = 3.52$, $df = 2.70$, $p = 0.0457$; equivalent to $F(1, 2.70) = 12.41$). Differences in abundance among the remaining sampling sites were not statistically significant ($p > 0.05$).

3.2 MP size

In order to evaluate variation across the different particle-size classes, one-way ANOVA was performed for each size group (Fig. 4). Isolated MPs were then analyzed down to a minimum detectable size of ≥ 0.15 mm, determined by the lowest size of the sieve used for separation and particles smaller than this threshold were not quantified.

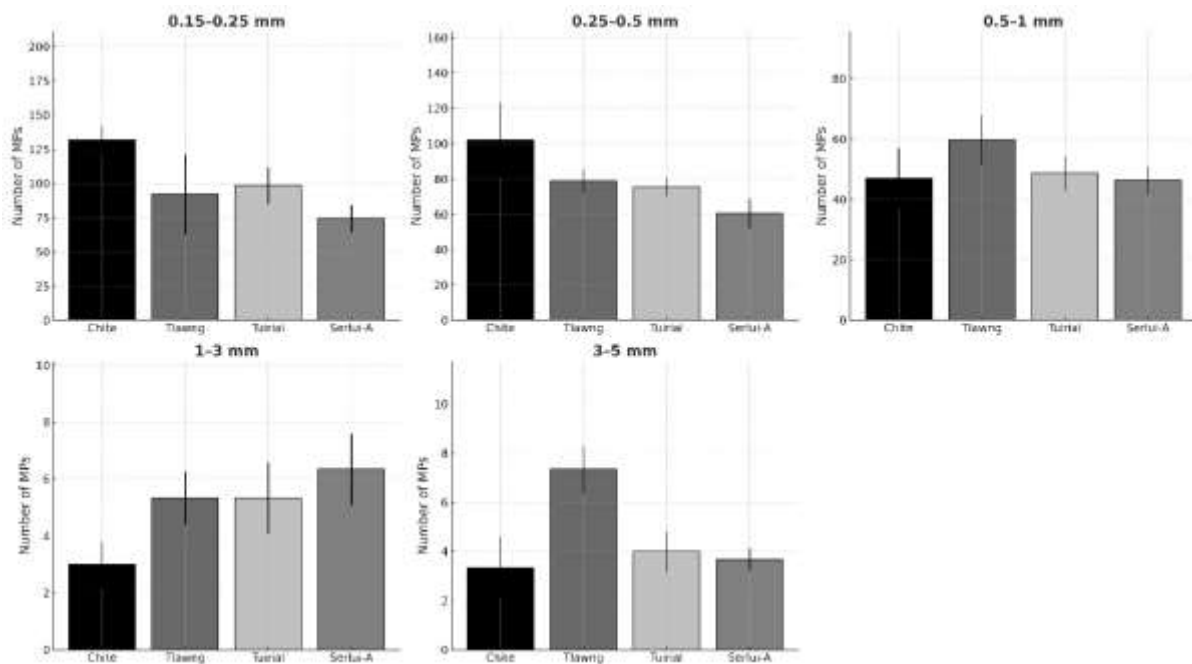


Fig. 4: Statistical analysis of abundance of different MP sizes collected from different sampling sites.

In addition, comparison of MP abundance of the different size groups across the four rivers revealed that the smallest 0.15–0.25 mm fraction displayed the highest mean counts in all sample sites, with the greatest abundance (131.67 particles) in Chite, followed by Tuirial (98.33), Tlawng (92.00), and Serlui-A (74.33). In terms of percentage, this size class accounted for 45.9% in Chite, 37.9% in Tlawng, 42.5% in Tuirial, and 38.9% in Serlui-A (Fig. 5). However, one-way ANOVA indicated that differences among rivers for this size class were not statistically significant ($p > 0.05$). The second most abundant fraction was the 0.25-0.5 mm group, contributing between 31–36%, with the highest mean (102.00 particles) present in Chite. No significant spatial variation was detected for this size fraction across the sites (one-way ANOVA, $p > 0.05$). The 0.5-1 mm group was found to be moderately abundant, ranging from 46–60 particles between the four rivers, contributing 16.4% (Chite), 24.6% (Tlawng), 21.0% (Tuirial), and 24.3% (Serlui-A) of total counts, with no statistically significant differences among rivers ($p > 0.05$). The larger groups (1-3 mm and 3–5 mm size) presented a very small proportion of total particles, typically below 5% of each river’s MP count, both contributing approximately between 1–3%, depending on the river. Significant differences among sampling sites were observed only for the 3–5 mm fraction as revealed by one-way ANOVA ($F(3, df_c) = F\text{-value}, p = 0.0079$). From these results, it can be observed that although overall MP loads differ between the sample sites of the four rivers, spatial variability is most pronounced in the larger-sized particles.

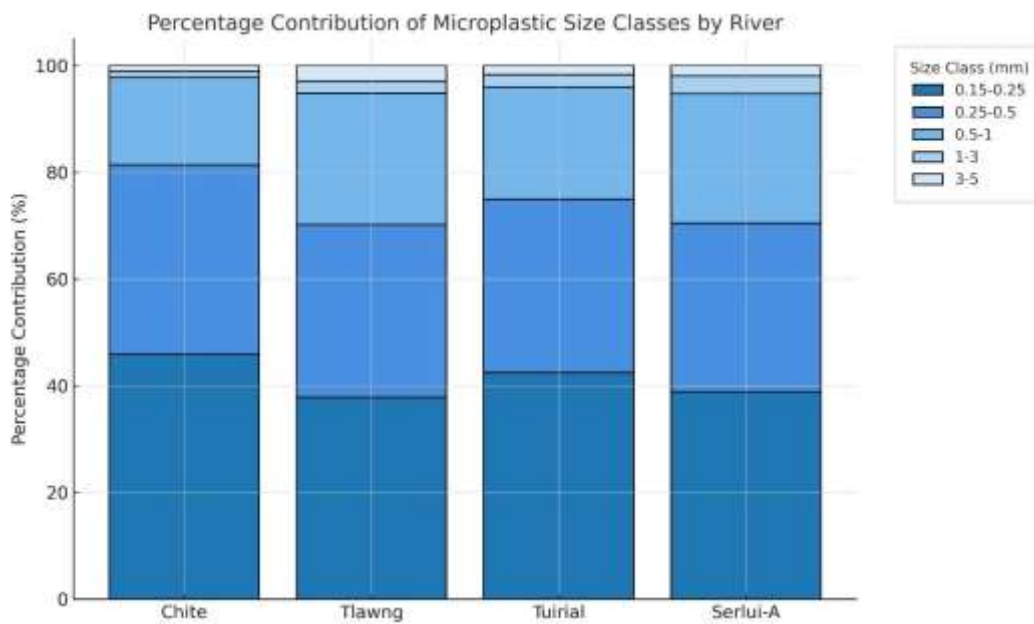


Fig. 5: Percentage contribution of MP size classes by river

Overall, these results determine a clear numerical dominance of smaller-sized MPs across all sampling sites, suggesting extensive fragmentation processes within these freshwater systems. However, statistically significant spatial differences were size-class dependent and restricted to the largest particle fraction, and therefore limits broader spatial interpretations.

3.3 MP shapes

MPs of varying shapes and structures were found in all the rivers. Fibers and fragment MPs were observed in all rivers. Fibers were the dominant shapes in all the sample sites accounting for approximately 40% of the total MPs recovered. Fragments and films together constituted approximately 45% of the particles, followed by spherical and foam structures (~10%). It also needs to be stated that shape-based categorization was derived from visual inspection, and therefore minor classification uncertainty cannot be excluded, particularly for irregular or weathered particles. Accordingly, the reported shape proportions need to be inferred as indicative trends rather than definitive morphological distributions.

The detailed distribution of various shapes identified from the samples is given in Fig. 6.

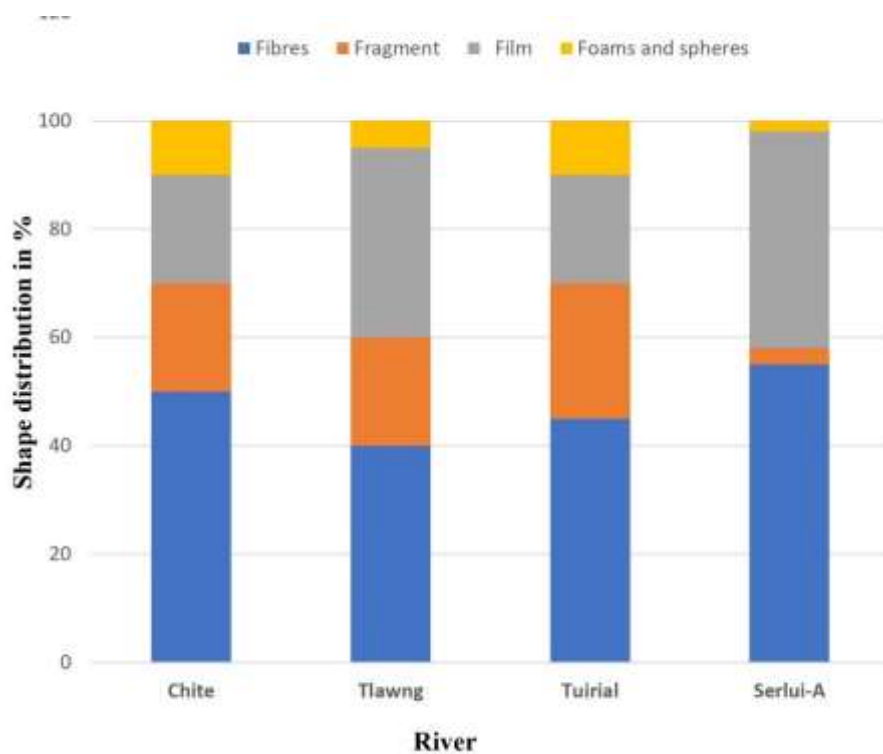


Fig. 6: Distribution of various MP shapes identified from sediment samples

The high abundance of fibre types in this study corroborates several other reports that have found MFs to be the dominant shape in fresh water bodies and sediments (Wang et al. 2017, Xu et al. 2021, Priya et al. 2023, Hosseinpour et al. 2025). Microscopic images of MPs comprising of different size ranges, shapes and colors are given in Fig. 7 and Fig. 8.

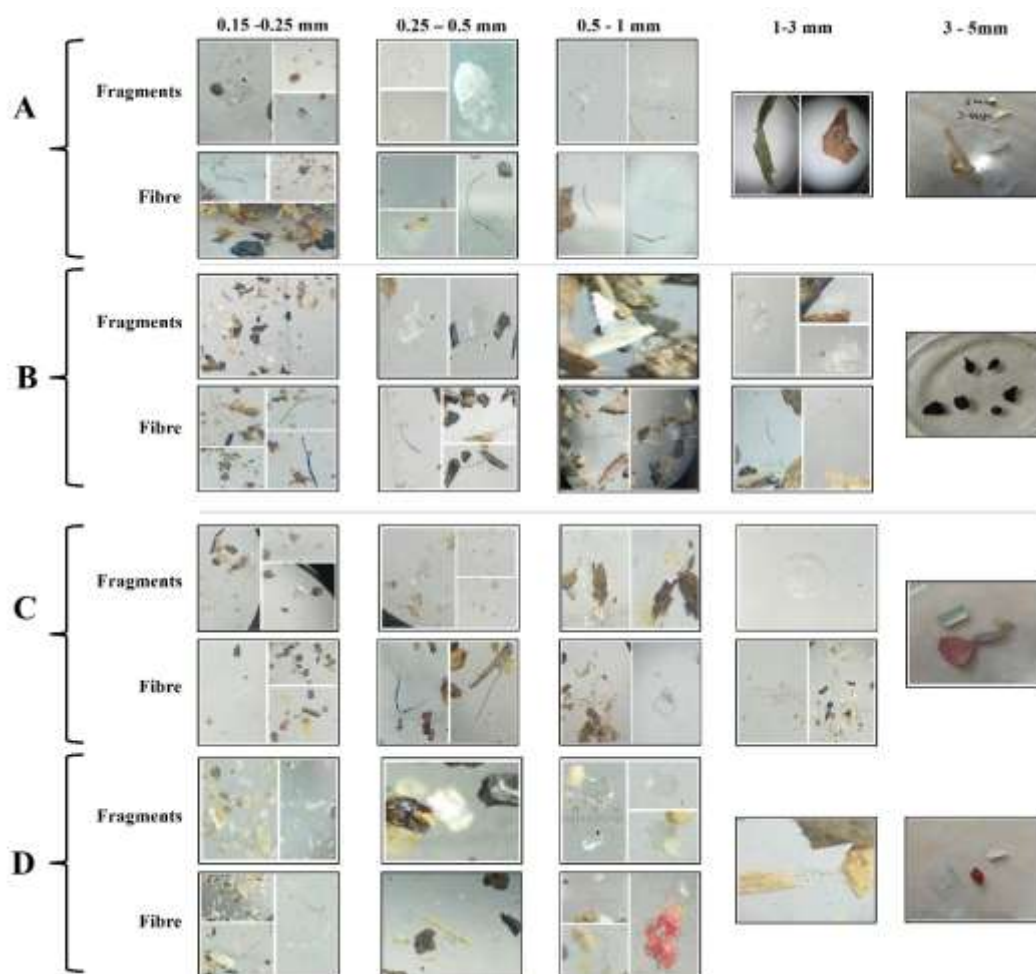


Fig. 7: Photographs of different MPs (fragments and fibres) isolated from sediment samples with their size under observed under a light microscope (10X and 40X magnifications). (A) MPs found in Chite river. (B) MPs found in Tlawng. (C) MPs found in Serlui-A. (D) MPs found in Tuirial.

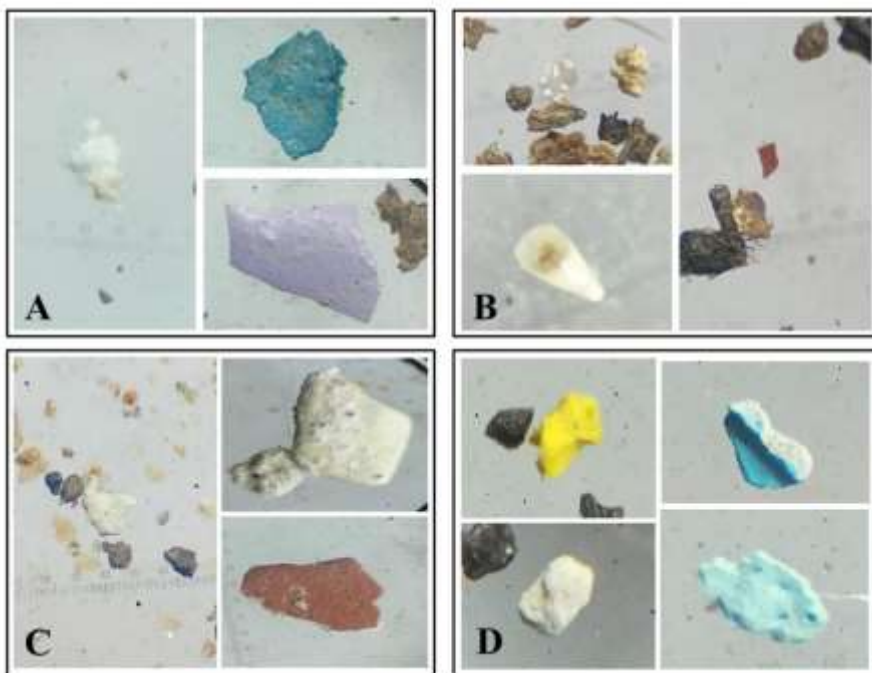


Fig. 8: Additional photographs of different MPs found in different rivers (A) MPs found in Chite river. (B) MPs found in Tlawng. (C) MPs found in Serlui-A. (D) MPs found in Tuirial.

3.4 Surface morphology and chemical characteristics

SEM analysis of different shapes of MPs (Fig. 9, A-D) showed variations in their surface morphologies, illustrating the heterogeneity in shape, texture, and surface characteristics typically associated with environmentally weathered polymers. The surfaces of fragments were observed to be uneven, rough and cracked, suggesting exposure to weathering, abrasion and disintegration over a period of time. Fig. 9-A, C, D show irregular, angular plastic fragment shapes with cracked surfaces and angular edges. Fig. 9-B displays intertwined, flexible fibre shape with smooth surfaces that are typically associated with anthropogenic textile sources.

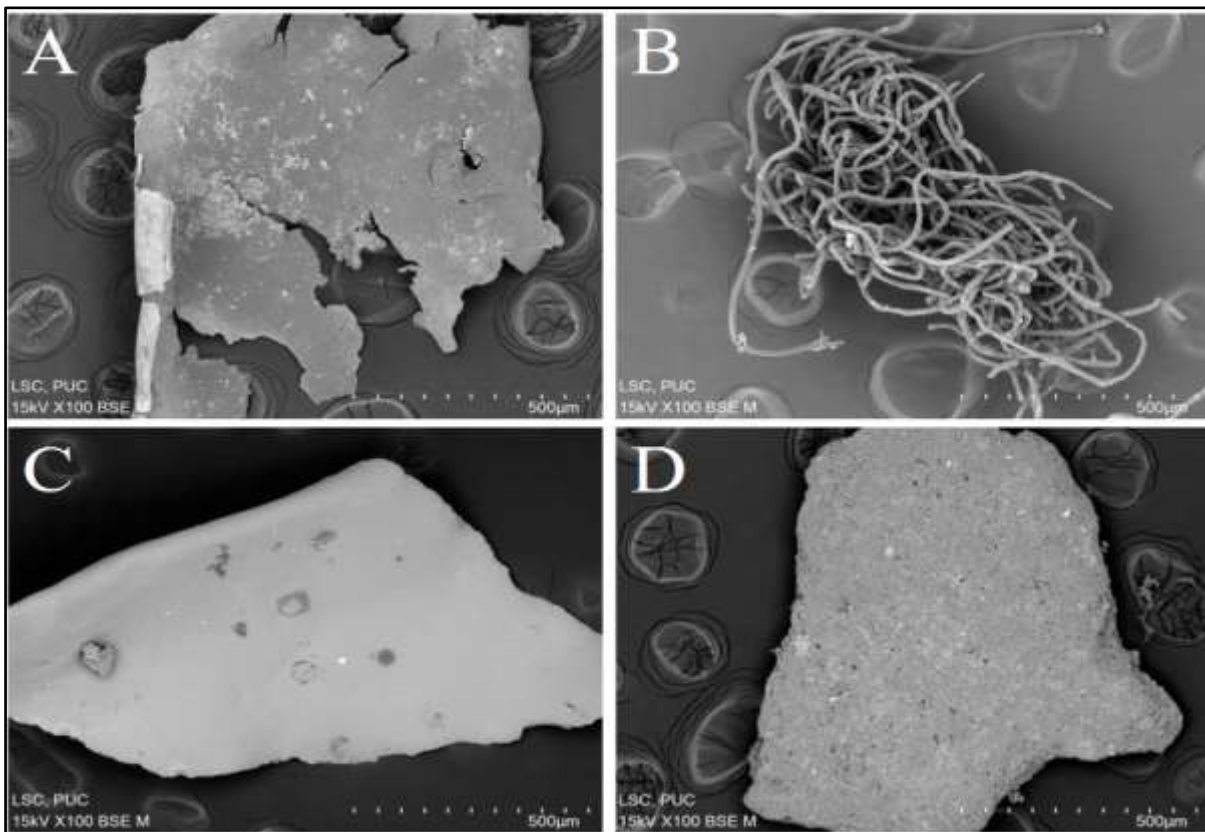


Fig. 9: SEM image of different MPs captured in 15kV at 100 magnifications. (A, C, D) Fragments (B) Fibres

FTIR analysis was done on 20 isolated particles (5 from each river within the 0.5-1mm size groups) that were randomly selected for polymer identification to validate visual identification. Spectral matching was conducted against a reference library, and particles were classified as MPs only when a match similarity $\geq 70\%$ was obtained, a threshold commonly adopted in environmental MP studies. Out of these, 75% (15 out of 20 particles) were thus confirmed to be MPs. Five samples could not be accurately identified due to inadequate spectrum intensity, that may imply a potential degree of overestimation associated with visual sorting alone. Therefore, polymer identification should be considered preliminary, and abundance values derived from visual inspection may include a limited proportion of non-plastic particles. The $\geq 70\%$ match similarity threshold was selected as per established MP FTIR studies, acknowledging that environmental degradation and surface fouling can lower spectral similarity scores without changing the underlying polymer identity. From this analysis, two distinct types of polymers namely, Polyethylene (PE) and Polypropylene (PP) were identified (Fig. 10), with PE being making up the majority of the polymers (93%) from the selected and analyzed samples. Spectra identifying PE showed two prominent aliphatic C–H stretching vibrations at $\sim 2916\text{ cm}^{-1}$ (asymmetric CH_2 stretch) and $\sim 2848\text{ cm}^{-1}$ (symmetric CH_2 stretch), accompanied by strong CH_2 deformation bands at $\sim 1470\text{ cm}^{-1}$ (scissoring) and $\sim 1463\text{ cm}^{-1}$ (bending). The most definitive marker of PE i.e., the CH_2 rocking vibration near $\sim 720\text{ cm}^{-1}$, was consistently present across all PE samples. The absence of carbonyl (C=O), aromatic (C=C), or halogen-containing (C–Cl) functional groups further supported the classification of these particles as PE, indicating a potential degree of overestimation associated with visual sorting alone. Therefore, polymer

identification in the present study should be considered preliminary, and abundance values derived from visual inspection may include a limited proportion of non-plastic particles.

Spectra identification for PP showed a strong CH_3 symmetric bending vibration at $\sim 1376 \text{ cm}^{-1}$, which serves as the primary indicator of PP. Additional CH_2 and CH_3 deformation modes were observed at $\sim 1455 \text{ cm}^{-1}$, along with C–H stretching vibrations at $\sim 2950\text{--}2838 \text{ cm}^{-1}$. The presence of skeletal C–C stretching and CH wagging modes in the $1165\text{--}997 \text{ cm}^{-1}$ region, together with an isotactic PP band near $\sim 840 \text{ cm}^{-1}$, further confirmed the identity of the PP polymer.

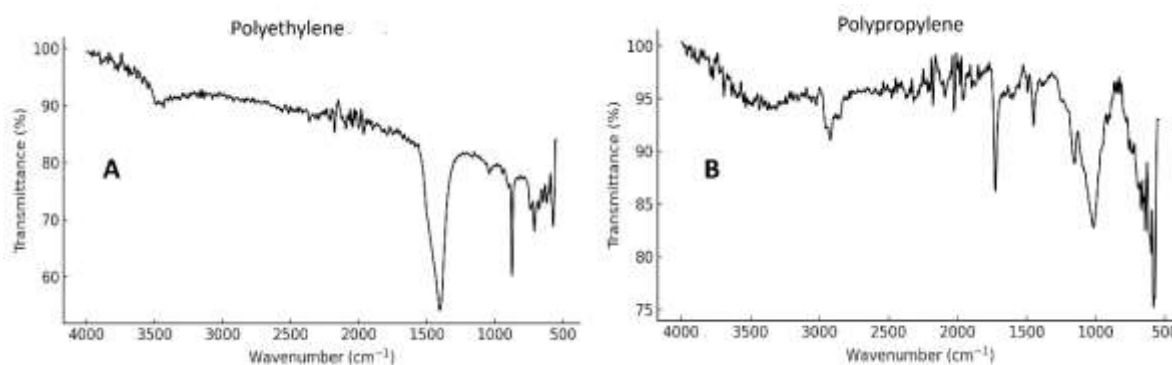


Fig. 10: FTIR spectra of MPs identified as Polyethylene (A) and Polypropylene (B)

4. DISCUSSION

This study aimed to evaluate the extent of MP pollution in shore sediments of rivers close to Aizawl city, Mizoram. The state lies in the north-eastern part of India and is situated in the lower eastern Himalayan region. Assessment results showed mean MP abundance to be 238.17 ± 46.95 particles/kg, and found to be highest in rivers that are relatively closer to Aizawl municipality, i.e., Chite river with the maximum number of MP particles, followed by Tlawng river. This corroborates various studies that have established positive correlation between elevated MP abundance and proximity of a freshwater body to urban and community areas (Peng et al., 2018, Tibbett et al. 2018, Kongkamee et al. 2024). However, it is critical to note that the interpretations presented in this study are site-specific rather than river-wide, as each river sediment was represented by a single sampling location and replicate samples ($n = 3$) constituted within-site subsamples as opposed to independent spatial replicates. Considering pronounced spatial heterogeneity characteristics for different riverine systems, the detected differences in MP abundance among the sampled locations should therefore be interpreted in light of the study's spatial limitations. Consequently, statistical comparisons among the four rivers also reflect relative differences only at the sampled sites and do not infer whole-river contamination patterns. While higher MP abundances were observed at sites in closer proximity to urbanized areas, these patterns should be considered indicative and exploratory, unless further established through multi-site, longitudinal, and seasonal sampling designs that can effectively resolve river-scale spatial variability. Additionally, mean abundance of MPs in shoreline sediments of other rivers in different parts of the Himalayas vary greatly. Table 2 gives a glimpse of the various studies that have been

conducted on MP pollution in different rivers within the Himalayan region and lists the mean abundance and types of polymers that have been detected. Values have been presented using original units (items/kg for sediments; items/L or items/m³ for water) from the published articles and categorized by sample matrix. The comparisons across studies are qualitative only, since the evident differences in units, sampling strategies, and analytical methods prevent direct numerical comparison.

Table 2: List of studies on MP pollution in sediments and water of other rivers of the Himalayan region, with mean abundances and polymer types

	Rivers	Sample	Abundance	Major MPs	Reference
Western Himalayan rivers	Indus	Sediment	525 - 1752 MP/kg	PP, PE, PES	Tsering et al. 2021
	Beas River	Water	46 - 222 items/L	PE	Bhaduri et al. 2025
		Sediment	36 - 896 items/kg		
	Chenab River	Water	34.66 - 45.98 MPs/m ³	PET, PP	Hafeez et al. 2023
	River Yamuna	Water	500 - 3900 MPs/m ³	PE, PS, PP	Vaid et al. 2022
	Ravi river	Water	768 - 1324 MPs/m ³	PP, PE, PES, and PS	Aslam et al. 2022
	Sutlej River	Water	3.6 - 19.0 MPs/L	PP, HDPE, PET	Nagendra et al. 2025
Soan River	Water	132.7 - 641.3 items/m ³	PET, PP, PE.	Jabeen et al. 2023	
Central Himalayan Rivers	River Ganga	Water	118.5 ± 49.65 particles /1000 L	PE, PP	Badola et al. 2023
		Sediment	131.5 ± 53.60 particles/kg		
	Upper Himalayan Ganga River	Water	100 - 1550 particles/L	PE, PA, PS	Chaudhary et al. 2025
		Sediment	50 - 1300 particles/Kg		
	River Ganga - (Suswa, Bindal and Rispana)	Water	2800 - 4200 items/L	PE, PP	Nayal & Suthar 2022
		Sediment	7200 - 16,400 items/kg		
	Karnali River	Water	0.12 - 0.64 particles/L	PE, PP	Maharjan et al. 2025
	Sapta-Gandaki river system	Water	24.7 - 61.2 particles/L	PE, PET, PVC	Kandel et al. 2025
Kosi river	Water	50 - 325 items/m ³	PE, PET, PA	Yang et al. 2021	
	Sediment	15 - 120 items/kg			
Eastern Himalayan Rivers	Brahmaputra	Sediment	531 - 3485 MP/kg	PP, PE, PA	Tsering et al. 2021
	Yarlung Tsangpo	Water	728.26 ± 100.53 items/m ³	PP, PE	Li et al. 2024
		Sediment	43.16 ± 5.82 items/kg		
	Lower Brahmaputra River	Water	11 - 32 MPs/L	Nylon, PET, PP	Hassan et al. 2025
	Manas river	Water	10 - 22 items/L	PP, PET, PS	Wang et al. 2021
	Water	22.35 - 27.94 MPs/L	PE, PP		

Jia Bharali River	Sediment	19.42 - 29 MPs/kg		Lahon & Handique 2024
Lasha river	Water	0.63 n/L	PP, PE	Zhou et al. 2023
	Sediment	0.37 n/g		
Kaljani River	Water	0.14 ± 0.11 pieces m ⁻³	PE, nylon, PP	Goswami et al. 2024
Karnaphuli River	Water	14.24 - 26.68 items m ⁻³	PET, HDPE, acrylonitrile butadiene styrene (ABS)	Jui et al. 2025

Unchecked pollution is still rampant in urban settlements, where domestic wastes are being discarded illicitly in side drains and open areas. Consequently, these are then carried off into rivers by wind and rainwater, and are further broken down into MPs via weathering. Although Tuirial river is the furthest in distance from Aizawl, MP counts were higher than Serlui-A which runs slightly closer to Aizawl. This could be attributed to its proximity to the rural settlement of Tuirial village around Tuirial river. As a result, anthropogenic activities such as mismanagement and dumping of garbage from the village either directly or indirectly into the river, ongoing construction of roads and a football field may have contributed to the elevated level of MPs in this river.

A considerable number of MPs were also detected in Serlui-A, which has various tributaries in the form of small streams, non-perennial brooks and gullies that originate from areas close to Aizawl municipality. Further, the presence of Li Cheng (or Dilmawi; a type of stream pool) which is a tourist attraction in Serlui-A could possibly be a key contributor of MPs in this river, due to indiscriminate disposal of plastic garbage by visitors. Although MP count in Serlui-A is the least when compared to other observed rivers, their detection in this river is particularly concerning because drinking water for Aizawl municipal areas is sourced from this river. However, since no studies on MP pollution in freshwater bodies within the state have been conducted as yet, conclusions cannot be made regarding the presence and abundance of MPs in the water, but further studies are tremendously warranted. Since Mizoram does not have large-scale manufacturing industries and complexes, the aggregation of MPs in the shore sediments of these rivers can mostly be attributed to inadequate domestic waste management procedures in the urban areas, and their subsequent runoff into surrounding water bodies and non-target areas due to the hilly terrain and high annual rainfall (Singh et al. 2023). These garbage masses are then evidently weathered down and fragmented into miniscule particles by various natural elements and environmental factors. Sediments eventually act as sinks for MP deposition, with gradual aggregation over time since they can easily be carried over long distances due to their miniscule sizes and lower densities (Ismanto et al. 2023, Napper et al. 2023).

The study found a higher abundance of smaller MPs below 1mm, especially within the size ranges of 0.15-0.25mm and 0.25-0.5mm in all four sampling locations while those above 3mm in size were significantly lower. Fragmentation processes contribute significantly to the size distribution of MPs in aquatic systems and tend to settle more easily due to their reduced buoyancy and greater susceptibility to deposition under lower energy conditions. (Browne et al. 2011). Larger plastic items break down into smaller particles as a result of physical, chemical, and biological factors and this continued weathering may explain why smaller MPs are found in higher

concentrations in sediments (Wang et al. 2022, Lu et al. 2023). Research by Klein et al. (2015) on the Rhine River revealed that the majority of MPs in sediments were smaller than 1 mm, with finer-grained sediments harbouring the highest concentrations of MPs. Similarly, a review by Yang et al. (2020) observed that in river systems, smaller MPs are usually more abundant than larger ones in sediments, and MPs lower than 0.5mm in particular were especially prevalent in sediment samples, further supporting the notion that smaller particles are more likely to accumulate in sediments due to their lower transport potential and easier incorporation into sediment layers. Another study by Ramaremsa et al. (2022) also reported decrease in abundance with increase in MP sizes in sediments of Vaal River in South Africa. In addition, other factors may also contribute to the abundance of MPs in sediments. Waldschläger and Schüttrumpf (2019) demonstrated that hydrodynamic conditions such as current velocity and turbulence significantly affect the settling velocities of MPs. Smaller particles settle more quickly due to reduced drag and hydrodynamic resistance, accumulating in riverbeds, especially in regions of decreased water flow such as oxbow lakes and estuaries, inferring that the smaller the particles, the greater the likelihood of their being present in river sediments. Although numerical differences in MP abundance were observed among rivers, particularly higher counts in Chite compared to the other sites, most size classes did not exhibit statistically significant spatial variation, which may suggest a broadly comparable distribution of smaller MPs across the river network. Statistical analyses demonstrated that spatial differences were size-dependent, with significant variation detected only in the largest particle fraction (3–5 mm). These results imply that larger particles may be more strongly influenced by localized sources, limited transport potential, and site-specific retention mechanisms, whereas smaller particles are more uniformly dispersed due to enhanced mobility and fragmentation. Since the analytical detection limit of this study was limited to ≥ 0.15 mm, MPs smaller than this threshold were not captured, and the reported abundances potentially underestimate the contribution of smaller size fractions, particularly fine fragments and fibres below 0.15 mm.

Fibres represented the majority of MP particles in all the rivers under study, constituting more than 40% of all MPs isolated. This corroborated other studies on freshwater bodies in other Indian river systems where MFs were the dominant type (Kalangutkar et al. 2023, Napper et al. 2023) as well as sediments of several rivers in other parts of the globe (Huang et al. 2021, Akdogan et al. 2023). Domestic and commercial laundering of various types of textiles have been known to be significant contributors of MFs in different aquatic bodies and is also a consequence of proximity to urban areas (Hazlehurst et al. 2023). Therefore, household wastes from residents of the city including effluents from washing textiles have possibly added to the abundance of MFs in the observed river sediments. Blue, red, and transparent fibres were the main MFs found in the sediment samples. The transparent fibres may indicate that fishing does contribute in some measure to the abundance of MFs in these rivers as net fishing is common in some parts of the selected rivers, especially Tlawng. However, given that fibres constituted a significant proportion of the detected MPs, absence of procedural blanks is recognized as a limitation of the present study. Since fibres can be particularly sensitive to airborne contamination during sampling and laboratory processing, stringent precautions were applied to the best extent possible during sample collection, handling, and analysis to minimize external contamination. In the absence of field or laboratory blanks, a quantitative assessment

of background contamination was not feasible, and therefore the reported fibre abundances should be appropriately interpreted, as minor contributions from secondary contamination cannot be fully ruled out.

Of the 20 particles selected for FTIR analysis from the visually identified MPs, 15 particles (75%) were spectroscopically confirmed as plastic polymers, while the remaining five particles could not be reliably identified due to insufficient spectral quality and were therefore, excluded from polymer classification. Consequently, polymer composition results are based solely on this subset of confirmed particles. Although visual microscopy remains a widely applied screening approach in MP studies, particularly for larger size fractions, the absence of auxiliary confirmation techniques such as hot needle testing, Nile Red staining, etc., represents a limitation of the present study and as a result, classification uncertainty cannot be fully excluded. Nevertheless, PE was found to be the dominant type of polymer, which underscores the pervasive use of PE-based materials in the region under study. PE is the primary constituent of various plastic products such as single-use packaging films, carry bags, wrappers, and household consumables, all of which are extensively used and frequently mismanaged in urban and peri-urban areas (Schwab et al. 2024). Our findings substantiate several other studies in different rivers (Doyen et al. 2019, Maheswaran et al. 2022, Babkiewicz et al. 2025) and also within the Himalayan region where PE has also been found to be one of the most abundant pollutant particles (Table 2). Comparatively, PP particles were significantly lower amongst the analyzed particles, which may be consistent with its use in more rigid consumer products such as bottle caps, storage containers, and packaging components, and its slightly higher density and mechanical strength likely reduces fragmentation rate relative to PE, possibly contributing to its lower representation in sediments (Khoironi et al. 2020, Anak Alexander Tampan et al. 2022). However, given the limited number of particles subjected to FTIR analysis, the polymer identification presented in this study is preliminary and indicative, intended to provide an overview of dominant polymer types rather than a complete characterization of sediment polymer composition. Additionally, although a ZnCl_2 density of 1.5 g cm^{-3} used in the isolation method enables effective recovery of most environmentally relevant polymers, it is acknowledged that some high-density polymers (e.g., PVC and highly compact or mineral-encrusted PET particles) may not be fully recovered, particularly when aging, biofouling, or aggregation can increase their effective density. Despite the use of two sequential extraction cycles, a degree of underestimation of high-density MPs cannot be entirely excluded. Hence, the reported polymer composition should therefore be interpreted as conservative with respect to dense polymer fractions.

5. CONCLUSIONS

This study provides a preliminary and baseline assessment of MP contamination in shoreline sediments of rivers that run close to Aizawl city, the highest urbanized region in the state of Mizoram. Importantly, this study presents one of the first systematic efforts to document MP contamination in site-specific freshwater riverine sediments from this part of the Eastern Himalayan region. Baseline information on MPs pollution in Mizoram has been virtually absent, despite the region being recognized as a biodiversity hotspot with ecologically sensitive river systems. By providing the first empirical evidence of MP occurrence in riverbank sediments in and close to Aizawl, the present work fills a critical knowledge gap and establishes a reference point for future monitoring and

comparative assessments. Such baseline data are essential for the early detection of emerging pollutants, formulation of region-specific management strategies, and prevention of further degradation of freshwater ecosystems in this fragile Himalayan landscape. Within the constraints of the study design, sampling coverage, and analytical resolution, this study also paints a harrowing picture of the extensive use and improper management of plastic products in Aizawl at large, since these are evidently polluting the rivers in close proximity to urban areas and settlements. More in-depth research concerning MP pollution is crucial to detect the presence and extent of contamination in air, soil, food and other river waters within the state since they are also susceptible to pollution from source to distant sites via river systems. Although lack of funding for this study has limited our preliminary investigation in terms of area of study, sample type and chemical analysis of particles, further studies on concentrations in river water, and ingestion by aquatic organisms are still lacking and are of utmost importance. In particular, tap water from Serlui-A needs further investigation for MP presence since it is a critical lifeline for thousands of people. However, the present study did not analyze drinking water samples and does not evaluate human exposure or health risk. Moreover, according to the World Health Organization, there are currently no universally accepted numeric safety thresholds for MPs in water, and regulatory and scientific bodies recommend enhanced monitoring and further toxicological research before such standards can be established (WHO, 2019). Nevertheless, the detected occurrence of MPs in riverine environments highlights the need for future integrated studies that include drinking water analysis, exposure assessment, and toxicological evaluation to better understand potential human health implications. Such investigations would be crucial for translating such findings into meaningful risk assessments.

Furthermore, there is a need for strengthened management strategies to limit the extensive use and disposal of plastics to minimize the indiscriminate pollution of our precious rivers, not just for the sake of humans, but all organisms associated with these aquatic environments. Single-use plastic bags, bottles, and packaging items should be ideally substituted by natural fibre products that are produced from bamboo, jute, cotton, etc. Although plastic pollution is a global issue, the state of Mizoram also still has a long way ahead in terms of combatting it, with an urgent need to provide alternatives to single-use plastics, and more efficient facilities for plastic recycling. In addition, collective awareness amongst the people needs to be inculcated regarding the harmful consequences of pollution on the environment and importance of mitigating pollution as a whole.

Abbreviations:

- ABS: acrylonitrile butadiene styrene
- HD/LD-PE: high-and low-density polyethylene
- MFs: Microplastic fibers
- MPs: Microplastics
- PA: Polyamide
- PE: Polyethylene
- PES: Polyester
- PET: Polyethylene terephthalate
- PS: Polystyrene

PP: Polypropylene

PVC: polyvinyl chloride

SEM: Scanning Electron Microscope

Author Contributions:

Joseph Vanlalsawma Sailo: Data curation, Writing- Original draft preparation, Investigation

Malsawmhriatzuala Jeremy: Writing- review & editing, Visualization.

Laldinsangi Chawngte: Conceptualization, Methodology, Supervision, Writing – review & editing.

Funding: This research received no external funding

Acknowledgments: The authors gratefully acknowledge the faculty of School of Life Sciences and the Research and Instrumentation Laboratory, Pachhunga University College for providing all the necessary facilities and instruments required to carry out this work.

Conflicts of Interest: The authors declare no conflicts of interest.

REFERENCES

1. Akdogan, Z., Guven, B. and Kideys, A.E., 2023. Microplastic distribution in the surface water and sediment of the Ergene River. *Environmental Research*, 234, p.116500. [DOI] [Google Scholar] [PubMed] [URL]
2. Viswanathan, P.M., 2022. Occurrence, distribution and sources of microplastics in beach sediments of Miri coast, NW Borneo. *Chemosphere*, 305, p.135368. [DOI] [Google Scholar] [PubMed] [URL]
3. Anbumani, S. and Kakkar, P., 2018. Ecotoxicological effects of microplastics on biota: a review. *Environmental Science and Pollution Research*, 25(15), pp.14373-14396. [DOI] [Google Scholar] [PubMed] [URL]
4. Aslam, M., Qadir, A., Hafeez, S., Aslam, H.M.U. and Ahmad, S.R., 2022. Spatiotemporal dynamics of microplastics burden in River Ravi, Pakistan. *Journal of Environmental Chemical Engineering*, 10(3), p.107652. [DOI] [Google Scholar]
5. Babkiewicz, E., Vecmane, E., Fuk, M., Jurgielewicz, M., Koniuk, A., Kurek, E., Maszczyk, P., Michalska-Kacymirow, M., Jonuskiene, D., Norvaišienė, J. and Burdukovska, V., 2025. Microplastics in the Baltic Sea region lakes—standardized insights reveal urban shoreline as key driver. *Environmental Science and Pollution Research*, 32(47), pp.27052-27067. [DOI] [Google Scholar] [URL]
6. Badola, N., Sobhan, F. and Chauhan, J.S., 2023. Microplastics in the River Ganga and its fishes: Study of a Himalayan River. *Science of The Total Environment*, 901, p.165924. [DOI] [Google Scholar] [PubMed] [URL]
7. Bajt, O., 2021. From plastics to microplastics and organisms. *FEBS Open bio*, 11(4), pp.954-966. [DOI] [Google Scholar] [PubMed] [URL]

8. Balestra, V. and Bellopede, R., 2022. Microplastic pollution in show cave sediments: First evidence and detection technique. *Environmental pollution*, 292, p.118261. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
9. Bhaduri, R.N., Sinha, S., Guerro, A.M., Jackson, S.L., Alemán, E.A. and Chatterjee, S., 2025. Microplastic contamination and environmental risks in the Beas River, western Himalayas. *Environmental Pollution*, 365, p.125387.. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
10. Bridson, J.H., Abbel, R., Smith, D.A., Northcott, G.L. and Gaw, S., 2023. Solving a microplastic dilemma? Evaluating additive release with a dynamic leaching method for microplastic assessment (DyLeMMA). *MethodsX*, 10, p.102221. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
11. Chaudhary, M., Rawat, S. and Suthar, S., 2025. Microplastic in upper Himalayan Ganga river: Occurrence, seasonal dynamics and ecological risk. *Science of The Total Environment*, 967, p.178824. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
12. Cole, M., Webb, H., Lindeque, P.K., Fileman, E.S., Halsband, C. and Galloway, T.S., 2014. Isolation of microplastics in biota-rich seawater samples and marine organisms. *Scientific reports*, 4(1), p.4528. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
13. Desforges, J.P.W., Galbraith, M., Dangerfield, N. and Ross, P.S., 2014. Widespread distribution of microplastics in subsurface seawater in the NE Pacific Ocean. *Marine pollution bulletin*, 79(1-2), pp.94-99. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
14. Doyen, P., Hermabessiere, L., Dehaut, A., Himber, C., Decodts, M., Degraeve, T., Delord, L., Gaboriaud, M., Moné, P., Sacco, J. and Tavernier, E., 2019. Occurrence and identification of microplastics in beach sediments from the Hauts-de-France region. *Environmental Science and Pollution Research*, 26(27), pp.28010-28021. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
15. Ghanbari, N., Fataei, E., Najj, A., Imani, A.A. and Nasehi, F., 2022. Microplastic pollution in sediments in the urban section of the Qara Su River, Iran. *Applied Water Science*, 12(8), p.192. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)
16. Ghosh, S., Sinha, J.K., Ghosh, S., Vashisth, K., Han, S. and Bhaskar, R., 2023. Microplastics as an emerging threat to the global environment and human health. *Sustainability*, 15(14), p.10821. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)
17. Goswami, P. and Bhadury, P., 2024. Characteristics of microplastics in tributaries of the upper Brahmaputra River along the Himalayan foothills, India. *Environmental Research Communications*, 6(7), p.075013. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)

18. Goyal, T., Singh, S., Das Gupta, G. and Verma, S.K., 2023. Microplastics in environment: a comprehension on sources, analytical detection, health concerns, and remediation. *Environmental Science and Pollution Research*, 30(54), pp.114707-114721. [DOI] [Google Scholar] [PubMed] [URL]
19. Gulizia, A.M., Philippa, B., Zacharuk, J., Motti, C.A. and Vamvounis, G., 2023. Plasticiser leaching from polyvinyl chloride microplastics and the implications for environmental risk assessment. *Marine Pollution Bulletin*, 195, p.115392. [DOI] [Google Scholar] [PubMed] [URL]
20. Gunaalan, K., Nielsen, T.G., Rodríguez Torres, R., Lorenz, C., Vianello, A., Andersen, C.A., Vollertsen, J. and Almeda, R., 2023. Is zooplankton an entry point of microplastics into the marine food web?. *Environmental science & technology*, 57(31), pp.11643-11655. [DOI] [Google Scholar] [PubMed] [URL]
21. Hafeez, S., Qadir, A., Aslam, M., Aslam, H.M.U., Rehmat, M.S. and Ahmad, S.R., 2023. Environmental risks of microplastics on the spatial and temporal gradient in a river originating from the western Himalayas. *Environmental Toxicology and Chemistry*, 42(3), pp.727-739. [DOI] [Google Scholar] [PubMed] [URL]
22. Hassan, M.A., Shammi, M. and Tareq, S.M., 2025. Assessing year-round microplastic loading in the lower Brahmaputra River: A threat to aquatic environment. *Journal of Hazardous Materials Advances*, 17, p.100592. [DOI] [Google Scholar] [URL]
23. Hazlehurst, A., Tiffin, L., Sumner, M. and Taylor, M., 2023. Quantification of microfibre release from textiles during domestic laundering. *Environmental Science and Pollution Research*, 30(15), pp.43932-43949. [DOI] [Google Scholar] [PubMed] [URL]
24. Hosseinpour, S., Keshavarzi, B., Moore, F. and Busquets, R., 2025. Microplastic pollution in playas—endorheic basins with closed drainage systems. A study in surface sediments of Bakhtegan–Tashk Lakes (South Iran). *Environmental Pollution*, p.127282. [DOI] [Google Scholar] [PubMed] [URL]
25. Huang, D., Li, X., Ouyang, Z., Zhao, X., Wu, R., Zhang, C., Lin, C., Li, Y. and Guo, X., 2021. The occurrence and abundance of microplastics in surface water and sediment of the West River downstream, in the south of China. *Science of the Total Environment*, 756, p.143857. [DOI] [Google Scholar] [PubMed] [URL]
26. Ismanto, A., Hadibarata, T., Sugianto, D.N., Zainuri, M., Kristanti, R.A., Wisha, U.J., Hernawan, U., Anindita, M.A., Gonsilou, A.P., Elshikh, M.S. and Al-Mohaimed, A.M., 2023. First evidence of microplastics in the water and sediment of Surakarta city river basin, Indonesia. *Marine Pollution Bulletin*, 196, p.115677. [DOI] [Google Scholar] [PubMed] [URL]

27. Jabeen, K., Xu, J., Liu, K., Zhu, L. and Li, D., 2023. Monthly variation and transport of microplastics from the Soan River into the Indus River. *Science of the Total Environment*, 905, p.166877. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
28. Jui, M., Miah, M.S., Islam, M.H., Sarwar, M.I., Moniruzzaman, M., Ankhy, R.S., Suchi, P.D. and Islam, M.S., 2025. Microplastic pollution in the water and sediment of the Karnaphuli River, Bangladesh: An ecological risk assessment. *Marine Pollution Bulletin*, 216, p.117948. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
29. Kalangutkar, N., Mhapsekar, S., Redkar, P., Valsan, G. and Warriar, A.K., 2024. Microplastic pollution in the Chapora River, Goa, Southwest India: Spatial distribution and risk assessment. *Environmental Monitoring and Assessment*, 196(5), p.409. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
30. Kandel, B., Adhikari, N., Chetri, A.K., Karki, A., Paudyal, H., Sharma, K.R., Giri, B. and Neupane, B.B., 2025. Distribution of microplastic contamination in Sapta-Gandaki river system, Nepal. *International Journal of Environmental Science and Technology*, 22(8), pp.7065-7076. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)
31. Karing, D.J., Anggiani, M., Cao, L.T.T. and El-shaammari, M., 2023. Occurrence of Microplastics in Kemena River and Niah River of Sarawak, Malaysia. *Tropical Environment, Biology, and Technology*, 1(1), pp.1-13. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)
32. Karthikeyan, P. and Subagunasekar, M., 2023. Microplastics pollution studies in India: A recent review of sources, abundances and research perspectives. *Regional Studies in Marine Science*, 61, p.102863. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)
33. Khoironi, A., Hadiyanto, H., Anggoro, S. and Sudarno, S., 2020. Evaluation of polypropylene plastic degradation and microplastic identification in sediments at Tambak Lorok coastal area, Semarang, Indonesia. *Marine pollution bulletin*, 151, p.110868. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
34. Klein, S., Worch, E. and Knepper, T.P., 2015. Occurrence and spatial distribution of microplastics in river shore sediments of the Rhine-Main area in Germany. *Environmental science & technology*, 49(10), pp.6070-6076. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
35. Koelmans, A.A., Nor, N.H.M., Hermsen, E., Kooi, M., Mintenig, S.M. and De France, J., 2019. Microplastics in freshwaters and drinking water: Critical review and assessment of data quality. *Water research*, 155, pp.410-422. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
36. Kongkamee, K., Chaimee, A., Hinhumpatch, P., Kitreerawutiwong, N., Keanjoom, R. and Yasaka, S., 2024. Microplastic contamination in rivers: a survey from the Nan River, Thailand. *The Lancet Planetary Health*, 8, p.S19. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)

-
37. Krause, S., Baranov, V., Nel, H.A., Drummond, J.D., Kukkola, A., Hoellein, T., Smith, G.H.S., Lewandowski, J., Bonet, B., Packman, A.I. and Sadler, J., 2021. Gathering at the top? Environmental controls of microplastic uptake and biomagnification in freshwater food webs. *Environmental Pollution*, 268, p.115750. [DOI] [Google Scholar] [PubMed] [URL]
38. Lahon, J. and Handique, S., 2024. Flood-induced variation and source apportionment of microplastics in Jia Bharali River of mid-Brahmaputra Valley, India. *Environmental Monitoring and Assessment*, 196(12), p.1236. [DOI] [Google Scholar] [PubMed] [URL]
39. Li, H., Lu, H., Feng, S., Xue, Y., Sun, T., Yan, Y., Zhang, X. and Yan, P., 2024. Environmental fate of microplastics in high-altitude basins: the insights into the Yarlung Tsangpo River Basin. *Journal of Environmental Management*, 365, p.121623. [DOI] [Google Scholar] [PubMed] [URL]
40. Lu, Q., Zhou, Y., Sui, Q. and Zhou, Y., 2023. Mechanism and characterization of microplastic aging process: A review. *Frontiers of Environmental Science & Engineering*, 17(8), p.100. [DOI] [Google Scholar] [PubMed] [URL]
41. Maharjan, K.K., Pyakurel, P., Bista, S. and Dhungel, R.P., 2025. Assessment of microplastic contamination in the Karnali River: A baseline study in remote region of Western Nepal. *Results in Engineering*, p.106562. [DOI] [Google Scholar] [URL]
42. Maheswaran, B., Karmegam, N., Al-Ansari, M., Subbaiya, R., Al-Humaid, L., Raj, J.S. and Govarthanam, M., 2022. Assessment, characterization, and quantification of microplastics from river sediments. *Chemosphere*, 298, p.134268. [DOI] [Google Scholar] [PubMed] [URL]
43. Masura, J., Baker, J., Foster, G. and Arthur, C., 2015. Laboratory methods for the analysis of microplastics in the marine environment: Recommendations for quantifying synthetic particles in waters and sediments. [Google Scholar] [URL]
44. McCormick, A.R., Hoellein, T.J., London, M.G., Hittie, J., Scott, J.W. and Kelly, J.J., 2016. Microplastic in surface waters of urban rivers: concentration, sources, and associated bacterial assemblages. *Ecosphere*, 7(11), p.e01556. [DOI] [Google Scholar] [URL]
45. Nagendra, P.S., Datta, S.N., Pathak, D., Tewari, G., Gupta, K. and Leishangthem, G.D., 2025. Assessment of Microplastic Contamination and Ecological Risk in the Middle Stretch of the Sutlej River, Punjab, India. *Asian Journal of Environment & Ecology*, 24(12), pp.33-51. [DOI] [Google Scholar] [URL]
46. Napper, I.E., Baroth, A., Barrett, A.C., Bhola, S., Chowdhury, G.W., Davies, B.F., Duncan, E.M., Kumar, S., Nelms, S.E., Niloy, M.N.H. and Nishat, B., 2023. The distribution and characterisation of microplastics in air, surface water and

- sediment within a major river system. *Science of the Total Environment*, 901, p.166640. [\[DOI\]](#) [\[Google Scholar\]](#)
[\[PubMed\]](#) [\[URL\]](#)
47. Nayal, R. and Suthar, S., 2022. First report on microplastics in tributaries of the upper Ganga River along Dehradun, India: Quantitative estimation and characterizations. *Journal of Hazardous Materials Advances*, 8, p.100190. [\[DOI\]](#)
[\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
48. Patil, P.B., Maity, S. and Sarkar, A., 2022. Potential human health risk assessment of microplastic exposure: current scenario and future perspectives. *Environmental monitoring and assessment*, 194(12), p.898. [\[DOI\]](#) [\[Google Scholar\]](#)
[\[PubMed\]](#) [\[URL\]](#)
49. Pautu, L., Lalmalsawma, P., Vanramliana, Balasubramani, K., Balabaskaran Nina, P., Rosangkima, G., Sarma, D.K., Malvi, Y. and Hunropuia, 2023. Seroprevalence of scrub typhus and other rickettsial diseases among the household rodents of Mizoram, North-East India. *Zoonoses and Public Health*, 70(3), pp.269-275. [\[DOI\]](#) [\[Google Scholar\]](#)
[\[PubMed\]](#) [\[URL\]](#)
50. Peng, G., Xu, P., Zhu, B., Bai, M. and Li, D., 2018. Microplastics in freshwater river sediments in Shanghai, China: a case study of risk assessment in mega-cities. *Environmental Pollution*, 234, pp.448-456. [\[DOI\]](#) [\[Google Scholar\]](#)
[\[PubMed\]](#) [\[URL\]](#)
51. Priya, K.K., Thilagam, H., Muthukumar, T., Gopalakrishnan, S. and Govarthanam, M., 2023. Impact of microfiber pollution on aquatic biota: A critical analysis of effects and preventive measures. *Science of the Total Environment*, 887, p.163984. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
52. Ramaremsa, G., Ndlovu, M. and Saad, D., 2022. Comparative assessment of microplastics in surface waters and sediments of the Vaal River, South Africa: abundance, composition, and sources. *Environmental Toxicology and Chemistry*, 41(12), pp.3029-3040. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
53. Rhodes, C.J., 2018. Plastic pollution and potential solutions. *Science progress*, 101(3), pp.207-260. [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
54. Saad, D., Ndlovu, M., Ramaremsa, G. and Tutu, H., 2022. Microplastics in freshwater environment: the first evaluation in sediment of the Vaal River, South Africa. *Heliyon*, 8(10). [\[DOI\]](#) [\[Google Scholar\]](#) [\[PubMed\]](#) [\[URL\]](#)
55. Sarijan, S., Azman, S., Said, M.I.M., Andu, Y. and Zon, N.F., 2018. Microplastics in sediment from Skudai and Tebrau river, Malaysia: a preliminary study. In *MATEC Web of Conferences* (Vol. 250, p. 06012). EDP Sciences. [\[DOI\]](#) [\[Google Scholar\]](#) [\[URL\]](#)

-
56. Schwab, S.T., Baur, M., Nelson, T.F. and Mecking, S., 2024. Synthesis and deconstruction of polyethylene-type materials. *Chemical Reviews*, 124(5), pp.2327-2351. [DOI] [Google Scholar] [PubMed] [URL]
57. Sendra, M., Pereiro, P., Figueras, A. and Novoa, B., 2021. An integrative toxicogenomic analysis of plastic additives. *Journal of Hazardous Materials*, 409, p.124975. [DOI] [Google Scholar] [PubMed] [URL]
58. Singh, A.P., Devi, A.S. and Sahoo, U.K., 2023. Quantifying plastic waste generation in Aizawl city, India: waste management and its impact on human and environmental health. *Environmental Science and Pollution Research*, 30(49), pp.107390-107402. [DOI] [Google Scholar] [PubMed] [URL]
59. Smith, M., Love, D.C., Rochman, C.M. and Neff, R.A., 2018. Microplastics in seafood and the implications for human health. *Current environmental health reports*, 5(3), pp.375-386. [DOI] [Google Scholar] [PubMed] [URL]
60. Tibbetts, J., Krause, S., Lynch, I. and Sambrook Smith, G.H., 2018. Abundance, distribution, and drivers of microplastic contamination in urban river environments. *Water*, 10(11), p.1597. [DOI] [Google Scholar] [URL]
61. Tsering, T., Sillanpää, M., Sillanpää, M., Viitala, M. and Reinikainen, S.P., 2021. Microplastics pollution in the Brahmaputra River and the Indus River of the Indian Himalaya. *Science of the Total Environment*, 789, p.147968. [DOI] [Google Scholar] [PubMed] [URL]
62. Vaid, M., Mehra, K., Sarma, K. and Gupta, A., 2022. Investigations on the co-occurrence of microplastics and other pollutants in the River Yamuna, Delhi. *Water Supply*, 22(12), pp.8767-8777. [DOI] [Google Scholar] [URL]
63. Van Cauwenberghe, L. and Janssen, C.R., 2014. Microplastics in bivalves cultured for human consumption. *Environmental pollution*, 193, pp.65-70. [DOI] [Google Scholar] [PubMed] [URL]
64. Vivekanand, A.C., Mohapatra, S. and Tyagi, V.K., 2021. Microplastics in aquatic environment: Challenges and perspectives. *Chemosphere*, 282, p.131151. [DOI] [Google Scholar] [PubMed] [URL]
65. Waldschläger, K. and Schüttrumpf, H., 2019. Effects of particle properties on the settling and rise velocities of microplastics in freshwater under laboratory conditions. *Environmental science & technology*, 53(4), pp.1958-1966. [DOI] [Google Scholar] [PubMed] [URL]
66. Wang, C., O'Connor, D., Wang, L., Wu, W.M., Luo, J. and Hou, D., 2022. Microplastics in urban runoff: Global occurrence and fate. *Water research*, 225, p.119129. [DOI] [Google Scholar] [PubMed] [URL]
67. Wang, G., Lu, J., Li, W., Ning, J., Zhou, L., Tong, Y., Liu, Z., Zhou, H. and Xiayihazi, N., 2021. Seasonal variation and risk assessment of microplastics in surface water of the Manas River Basin, China. *Ecotoxicology and environmental safety*, 208, p.111477. [DOI] [Google Scholar] [PubMed] [URL]

-
68. Wang, T., Li, B., Shi, H., Ding, Y., Chen, H., Yuan, F., Liu, R. and Zou, X., 2024. The processes and transport fluxes of land-based macroplastics and microplastics entering the ocean via rivers. *Journal of Hazardous Materials*, 466, p.133623. [DOI] [Google Scholar] [PubMed] [URL]
69. Wang, W., Ndungu, A.W., Li, Z. and Wang, J., 2017. Microplastics pollution in inland freshwaters of China: A case study in urban surface waters of Wuhan, China. *Science of the Total Environment*, 575, pp.1369-1374. [DOI] [Google Scholar] [PubMed] [URL]
70. World Health Organization, 2019. WHO calls for more research into microplastics and a crackdown on plastic pollution. *World Health Organization*. [Google Scholar] [URL]
71. Xu, Y., Chan, F.K.S., Stanton, T., Johnson, M.F., Kay, P., He, J., Wang, J., Kong, C., Wang, Z., Liu, D. and Xu, Y., 2021. Synthesis of dominant plastic microfibre prevalence and pollution control feasibility in Chinese freshwater environments. *Science of The Total Environment*, 783, p.146863. [DOI] [Google Scholar] [PubMed] [URL]
72. Yang, L., Luo, W., Zhao, P., Zhang, Y., Kang, S., Giesy, J.P. and Zhang, F., 2021. Microplastics in the Koshi River, a remote alpine river crossing the Himalayas from China to Nepal. *Environmental Pollution*, 290, p.118121. [DOI] [Google Scholar] [PubMed] [URL]
73. Yang, L., Zhang, Y., Kang, S., Wang, Z. and Wu, C., 2021. Microplastics in freshwater sediment: A review on methods, occurrence, and sources. *Science of the Total Environment*, 754, p.141948. [DOI] [Google Scholar] [PubMed] [URL]
74. Zhao, Q., Zhu, L., Weng, J., Jin, Z., Cao, Y., Jiang, H. and Zhang, Z., 2023. Detection and characterization of microplastics in the human testis and semen. *Science of The Total Environment*, 877, p.162713. [DOI] [Google Scholar] [PubMed] [URL]
75. Zhou, A., Zhao, Y., Liu, M., Suyamud, B., Yuan, W. and Yang, Y., 2023. Occurrence and risk assessment of microplastics in the Lhasa River—a remote plateau river on the Qinghai-Tibet Plateau, China. *Environmental Monitoring and Assessment*, 195(3), p.433. [DOI] [Google Scholar] [PubMed] [URL]